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Design and Synthesis of Fluorinated Iron Chelators for Metabolic Study and Brain Uptake

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(5) Supporting Information

ABSTRACT: A range of fluorinated 3-hydroxypyridin-4-ones has been synthesized where fluorine or fluorinated substituent was attached at 2- or 5- position of the pyridine ring in order to improve chemical and biological properties of 3hydroxypyridin-4-ones. The synthetic route is different from conventional counterparts where a functional group is introduced to a preformed 3-hydroxypyridin-4-one ring. Herein, we introduce a novel method which starts with a fluorine containing



precursor and the two hydroxyl groups at 3- and 4- positions of the pyridine ring are introduced at a later stage. The pK_3 values of the free ligands and the affinity constants of their iron complexes demonstrate that the presence of fluorine dramatically alters the values. The distribution coefficient values of the free ligands and corresponding iron(III) complexes between 1-octanol and MOPS buffer (pH 7.4) are also influenced. Glucuronidation and oxidation studies of selected fluorinated 3-hydroxypyridin-4ones demonstrate that some such fluorinated compounds have clear advantage over deferiprone in that they are metabolized more slowly. Blood-brain barrier permeability studies indicated that although lipophilicity influences the permeability it is not the only factor. Two of the selected seven fluorinated 3-hydroxypyridin-4-ones have improved brain distribution when compared with deferiprone.

INTRODUCTION

The presence of one or more fluorine atoms in drug candidates impacts a variety of properties on such molecules, including enhanced metabolic stability, bioavailability, selective reactivity, receptor binding interactions, and changes in physical properties.¹⁻⁴ Generally, the unique characterization of fluorinated molecules is attributed to the fluorine atom possessing a high electronegativity, relatively small size, increased thermal and oxidative stability, and altered molecular lipophilicity. The participation of fluorine in medicinal chemistry and drug design cannot be underestimated; in 2007, 115 fluorinated drug candidates were investigated in phase II studies and 44 in phase III clinical trials.⁵

There is an increasing interest in the application of ironselective chelators for the treatment and possible prevention of the onset of a range of neurodegenerative disorders, including Alzheimer's disease and Parkinson's disease.⁶ Indeed one such chelator, deferiprone (DFP), a 3-hydroxypyridin-4-one (HPO), is currently being used to treat Friedreich's ataxia (Chart 1). Although deferiprone is able to cross the blood-brain barrier, it is not particularly efficient at doing so.⁸ In addition, deferiprone

Chart 1. Structure of DFP and CP94



CP94: R=Et

is rapidly metabolized in the liver⁹ and thus the dose of deferiprone required to maintain a thalassaemia patient in negative iron balance is relatively high, typically in the region of 75 to 100 mg·kg⁻¹·day⁻¹.¹⁰ Another HPO analogue, 1,2diethyl-3-hydroxypyridin-4-one (CP94), has also been investigated.¹¹⁻¹³ Although a metabolic study in the rat demonstrated that CP94 retains the ability to chelate iron,¹⁴ a parallel study in man demonstrated that CP94 is rapidly converted to the 3-O-glucuronide conjugate.¹² In an attempt to improve the distribution properties of HPOs, we have investigated the influence of introducing fluorine atoms into the aromatic nucleus. In the present study, we have synthesized a series of monofluoro, difluoro, trifluoro, and trifluoromethyl substituted HPOs. The physicochemical properties of this series of compounds have been characterized, together with their metabolic stability and rates of brain uptake.

RESULTS AND DISCUSSION

Chemistry. There are two possible approaches to the introduction of fluorine into HPOs: one is to introduce fluorine or a fluorinated moiety into the preformed HPO matrix and the other is to start with a fluorine-containing precursor and introduce chelating functional groups. In this paper, we report three fluorinated HPOs synthesized by the former approach and 23 by the latter.

The synthetic route employed by the former approach is summarized in Scheme 1. 3,4-Bis(benzyloxy)pyridine-2-carbaldehyde 2 and 3-(benzyloxy)-2-(hydroxymethyl)-1,6-dimethyl-

Received: November 1, 2011 Published: February 16, 2012 Scheme 1. Synthesis of 2-Trifluoromethyl Substitution of 3-Hydroxypyridin-4-ones^a



^a(a) Me₃SiCF₃/TBAF; (b) DMSO/TEA/Py-SO₃; (c) Pd/C, H₂; (d) BCl₃.





^{*a*}(a) (i) LDA in THF at -75 °C for 0.5 h, (ii) B(OMe)₃ at -75 °C for 2 h, (iii) CH₃CO₃H at 0 °C for 1 h; (b) K₂CO₃, RI in acetone refluxed overnight; (c) (i) LTMP in THF at -75 °C for 1 h, (ii) B(OMe)₃ at -75 °C for 2 h, (iii) CH₃CO₃H at 0 °C for 1 h; (d) BBr₃ in DCM at 0 °C overnight; (e) (i) LDMEA in THF at -75 °C for 20 h, (ii) MeI; (f) (i) LTMP in THF at -75 °C for 20 h, (ii) MeI; (f) (i) LTMP in THF at -75 °C for 20 h, (ii) MeI; (f) (i) LTMP in THF at -75 °C for 20 h, (ii) MeI (PG = protected group).

pyridin-4-one **5** were readily prepared from commercially available maltol and kojic acid respectively by following a previous reported procedure.^{15,16} Selective oxidation of the alcohol **5** to its corresponding aldehyde **6** proceeded effectively

by using the sulfur trioxide pyridine complex $(SO_3 \cdot Py)$ in combination with dimethyl sulfoxide as the oxidizing reagent. The trifluoromethylation reaction was carried out by reacting the aldehyde with TMSCF₃ in the presence of a catalytic

Scheme 3. Synthesis of 5-Fluoro Substituted 3-Hydroxypyridin-4-one Derivatives^a



"(a) (i) LDA in THF at -75 °C for 2 h, (ii) B(OMe)₃ at -75 °C for 2 h, (iii) CH₃CO₃H at 0 °C for 1 h; (b) K₂CO₃, RI in acetone refluxed overnight; (c) (i) LTMP in THF at -75 °C for 20 h, (ii) B(OMe)₃ at -75 °C for 2 h, (iii) CH₃CO₃H at 0 °C for 1 h; (d) Pd(OH)₂/H₂/Et₃N; (e) (i) LTMP in THF at -75 °C for 24 h, (ii) MeI; (f) BBr₃ in DCM at 0 °C overnight; (g) RI in acetone, reflux overnight; (h) (i) LDA in THF at -75 °C for 20 h, (ii) MeI; (j) NaOMe, reflux overnight; (k) LDA in THF at -75 °C for 20 h, (ii) B(OMe)₃ at -75 °C for 2 h, (iii) CH₃CO₃H at 0 °C for 2 h, (iii) CH₃CO₃H at 0 °C for 1 h; (b) K₂CO₃H at 0 °C for 1 h; (c) LDA in THF at -75 °C for 20 h, (ii) MeI; (j) NaOMe, reflux overnight; (k) LDA in THF at -75 °C for 20 h, (ii) B(OMe)₃ at -75 °C for 2 h, (iii) CH₃CO₃H at 0 °C for 1 h (PG = protected group).

amount of tetrabutylammonium fluoride, followed by desilylation. The resulting alcohol was either hydrogenated to afford deprotected product **1a** or further oxidized to aldehydes **4** and **8**, respectively. As hydrogenation would reduce the ketone to its corresponding alcohol, boron trichloride was selected to remove the benzyl protecting group to form **1b** and **1c**, respectively.

We also investigated the ability of several fluoronitrogen agents such as Selectfluor to introduce fluorine directly into the heterocyclic ring, but unfortunately all such attempts failed. Therefore, the second approach which used commercial available fluorine-containing building blocks as starting materials was adopted.

The general methodology adopted for the synthesis of 2fluoro substituted HPOs 1d-m started from 2-fluoropyridine or 2-fluoro-5-methylpyridine is summarized in Scheme 2. 2-Fluoro-3-hydroxypyridine 10 was obtained by treating 2fluoropyridine 9 with lithium diisopropylamide (LDA), followed by successive additions of trimethylborate and peracetic acid. Lithiation occurs selectively at C3 of 2fluoropyridine due to a strong inductive effect of the fluorine atom.^{17,18} Before a second lithiation at C4 by using lithium 2,2,6,6-tetramethylpiperidine (LTMP), the 3-hydroxy group of the compound 10 required protection. Reacting 10 with methyl iodide in the presence of potassium carbonate affords a quantitative yield of 2-fluoro-3-methoxypyridine 11. A similar methodology was adopted to introduce a second hydroxyl group at C4 of compound 11. We attempted to protect the 4hydroxy group of 12 also using K₂CO₃/RI reagents. However, at this stage, a mixture of two alkylated products 13 and 14

were isolated, where the alkyl group was attached either on the oxygen of the 4-hydroxy or the nitrogen of the pyridine ring. The formation of 13 and 14 can be explained by the tautomerism of compound 12 between 4-hydroxypyridine and pyridine-4-one tautomers. By increasing the size of the alkyl group, the ratio of 13:14 can be dramatically increased. For example, a ratio of 4:5 for methyl group is enhanced to 30:1 for the isopropyl group. The 4-alkoxypyridine isomer 13 can be selectively methylated at either C5 or C6 to afford 16 and 15, respectively, depending on the choice of lithiation reagents. LTMP favors the deprotonation of the pyridine derivative at C5, while the unimetal superbase (USB: 1:1 ratio of butyllithium and lithium 2-(dimethylamino)ethoxide)^{17,19} prefers at C6. In contrast to the pyridine isomer 13, the pyridinone 14 cannot be methylated at either of the apparently vacant positions.

Compound 16 can also be produced from 2-fluoro-5methylpyridine 18, where only two alkoxy groups at both C3 and C4 are involved, in a similar way to that mentioned above. However, analogue 15 cannot be produced from the similar route, as 2-fluoro-6-methylpyridine is prone to be lithiated at the 6-methyl group in preference to C3. Compound 16 was again methylated by USB, followed by trapping with methyl iodide to afford the dimethyl substituted compound 17. Again, when compound 20 was refluxed with K_2CO_3/MeI in acetone, an equimolecular mixture of 16 and 21 was obtained. A conventional method to introduce an alkyl group on the N1 of pyridine ring using alkyl iodide or dimethyl sulfate to form 1alkylpyridinium failed with 2-fluoropyridine derivatives, most likely due to the strong influence of fluorine at C2. The alkyl Scheme 4. Synthesis of Multifluoro Substituted 3-Hydroxypyridin-4-ones^a



"(a) NaOMe; (b) Pd/C, HCOONH₄; (c) (i) LDA in THF at -75 °C for 0.5 h, (ii) B(OMe)₃ at -75 °C for 2 h, (iii) CH₃CO₃H at 0 °C for 1 h; (d) BBr₃, overnight.

Scheme 5. Synthesis of Trifluoromethylated 3-Hydroxypyridin-4-ones^a



a(a) (i) LTMP/16 h, (ii) B(OMe)₃/2 h, (iii) CH₃CO₃H/1 h; (b) CH₃I/K₂CO₃; (c) same as (a) except 3 h with LTMP; (d) BBr₃; (e) LTMP/MeI.

protecting group was then readily removed using BBr_3 to form the corresponding iron chelating products 1d-m.

The synthesis of 5-fluoro substituted HPOs 1n-u is outlined in Scheme 3. Although 3-fluoropyridine is a commercially available chemical and its lithiation by LDA occurs at C4 rather than C2, further lithiation readily lead to attacks at C2, as the proton at this position is more acidic than those at the 5- and 6positions due to the fluorine at C3. Therefore, 2-chloro-3fluoropyridine 22 was selected for the starting material, where the C2 was chlorine blocked. In similar fashion to that of 2fluoropyridine, the hydroxyl group was subsequently introduced at C4 and C5 to form intermediate 25. Methylation occurs at C6 when the 5-hydroxy group of 25 was protected to afford 27. The blocking chlorine atom of compounds 25, 26, and 27 was removed by hydrogenation to produce 30, 31, and 28, respectively. Compound 30 can be also obtained from commercially available 4-chloro-3-fluoropyridine 33 in a twostep manner via intermediate 34. Compound 31 can be obtained from 25 by either protection of 5-hydroxy group followed by reduction to remove the chlorine atom or a reverse procedure via 30. Because of the presence of a fluorine at C3, C2 of compound 31 prefer to be deprotonated over C6, followed by quenching by methyl iodide to obtain 32. In contrast to the 2-fluoropyridine analogues, which failed to produce 1-alkylpyridinium salts by reacting with alkyl iodide, when 3-fluoropyridine derivatives 28 and 30 were treated with alkyl iodide, a clear new spot at the bottom of TLC plate was detected. The resulting product was identified as the 1alkylpyridin-4-one derivatives 29 and 1r-u, respectively. A plausible mechanism for this unexpected product is explained by the formation of the unstable 1-alkylpyridinium salt intermediate.²⁰ 4- and 5-Alkyl protecting groups of 28-30 and 32 were removed by BBr₃ to afford the chelators 1n-q, respectively.

The syntheses of tri- and difluoro HPOs 1v and 1w started with the pentahalo substituted pyridine 35 (Scheme 4). Treatment of the commercially available 3-chloro-2,4,5,6tetrafluoropyridine or 3,5-dichloro-2,4,6-trifluoropyridine 35with 1 equiv of sodium methoxide yielded 36 in a good yield. Treatment of 36 with 10% Pd/C in the presence of ammonium formate at 50 °C for 10 h gave compound 37 with a high yield, followed by subsequent lithiation, electrophilic substitution, and oxidation as outlined above, introduced a hydroxyl group to afford compound 38. The 4-methyl protecting group was removed to produce 1v and 1w, respectively.

In similar fashion to that of 2-fluoro HPOs, the synthesis of 2-trifluoromethyl HPOs was initiated from 2-trifluoromethylpyridine 39 (Scheme 5). Because of the lower electron withdrawing influence of the trifluoromethyl group compared to fluorine at C2, deprotonation at C3 requires a stronger base, such as LTMP (pK_a 37.3), rather than LDA (pK_a 35.7), as a lithiation reagent. After two hydroxyl groups were introduced at C3 and C4 in a similar procedure as reported for that of 2fluoropyridine, compound 42 was treated with methyl iodide in the presence of K₂CO₃ to afford two methylated products 43 and 44 in a ratio of 3:2. Intermediate 43 was then methylated at C5 using LTMP/MeI reagents to obtain 45 in a high yield. Deprotection of compounds 42 and 44 with BBr₃ gave the expected products 1x and 1y, respectively. However, when 45 was treated with BBr₃ in same way, no peak was found in ¹⁹F NMR spectrum. Combined with the NMR, MS, and elemental analysis data, the product was deduced to be compound 1z. This result can be explained by the fact that the trifluoromethyl was substituted to tribromomethyl with BBr3 and then undergoes hydrolysis with methanol to form the ester (1z).

Determination of Acid Dissociation Constants. pK_a values of all the F-HPOs 1a-z are presented in Table 1. Similar to nonfluorinated analogues, all 1-alkyl substituted F-HPOs contain two pK_as , assigned to dissociable 3-hydroxy

Table 1. pK_as and Iron Affinities of Fluorinated 3-Hydroxypyridin-4-ones



compd	R ₁	R_2	R ₅	R ₆	pK_{a1}	pK _{a2}	pK _{a3}	$\log \beta_3$	pFe ³⁺
DFP	Me	Me	Н	Н	3.68	9.77		36.4	20.6
CP94	Et	Et	Н	Н	3.81	9.93		36.8	20.5
1a	Н	CH(OH)CF ₃	Н	Н	2.6	8.2		33.0	20.8
1b	Н	COCF ₃	Н	Н	2.8	6.4	9.7	28.6	19.0
1c	Me	COCF ₃	Н	Me	3.85	9.15		35.5	18.7
1d	Н	F	Н	Н	0.9	6.5	10.7	37-39 ^a	$18 - 20^{a}$
1e	Me	F	Н	Н	2.4	7.7		30.3	19.4
1f	Et	F	Н	Н	2.46	7.72		30.5	19.6
1g	<i>n</i> -Pr	F	Н	Н	2.49	7.89		30.7	19.5
1h	<i>i</i> -Pr	F	Н	Н	2.51	7.88		31.3	20.0
1i	<i>n</i> -Bu	F	Н	Н	2.61	7.79		31.2	20.1
1j	Н	F	Н	Me	1.15	6.7	11.08	37-39 ^a	17–19 ^a
1k	Н	F	Me	Н	1.42	6.66	11.10	37-39 ^a	16.5–18.5 ^{<i>a</i>}
11	Н	F	Me	Me	2.01	6.98	11.56	37-39 ^a	$16.5 - 18^{a}$
1m	Me	F	Me	Н	2.5	8.35		33.0	18.0
1n	Н	Me	F	Η	2.2	8.93	12.05	33.0	18.9
10	Me	Me	F	Н	2.24	9.08		33.0	18.5
1p	Н	Н	F	Me	1.94	8.68	12.04	32.0	18.6
1q	Н	Н	F	Н	1.6	8.2	11.7	29.9	17.8
1r	Me	Н	F	Η	1.76	8.18		30.9	18.9
1s	Et	Н	F	Η	1.78	8.25		31.9	19.7
1t	<i>n</i> -Pr	Н	F	Η	1.49	8.12		30.6	18.7
1u	<i>i</i> -Pr	Н	F	Η	1.72	8.12		29.7	17.9
1v	Н	F	F	F	<0.8	4.4	9.9	35.9	18.8
1w	Н	F	Η	F	<0.8	5.7	10.6	38.6	19.8
1x	Н	CF ₃	Η	Η	1.54	6.5	10.4	38.5-39.5 ^a	20-21 ^a
1y	Me	CF ₃	Н	Н	<0.8	7.4		26.5	16.2
1z	Н	COOMe	Me	Н	3.84	8.98		33.5	19.5
^a Range of va	lues quoted	due to the multiple	protonation	n of the iron	n complexes.				

Scheme 6. Proton Dissociation Equilibria of the 1-Nonsubstituted F-HPOs (Either R_2 or $R_5 = F$ or F-Containing Group) (For Clarity, the Resonance Forms Are Not Presented)



 (pK_{a2}) and 4-oxo group (pK_{a1}) .²¹ However, both $pK_{a}s$ are lower when compared to those of nonfluorinated HPOs, for example, compare **1e** with DFP. This is due to the strong electron withdrawing property of fluorine (Table 1). 1-Unsubstituted F-HPOs possess an additional pK_{a} which is

due to dissociation of 1-NH. This phenomenon does not occur on nonfluorinated analogues (1z) as the pK_a value falls outside the detectable range (pH 1–12).²² The ionization equilibrium of this type of F-HPOs is shown in Scheme 6. According to our observations on the absorption spectra shifts with increased pH

Table 2. Comparison of Distribution Coefficient of Free Ligand and Iron Complex



compd	R_1	R ₂	R ₅	R ₆	$\log D_{7.4}$ (n = 7) (free ligand)	$F_{\rm n}$	log P (free ligand)	$\log D_{7.4} (n = 4)^a (\text{Fe}^{3+} \text{ complex})$
DFP	Me	Me	Н	Н	-0.77 ± 0.02	0.995	-0.77	-2.60 ± 0.05
CP94	Et	Et	Н	Н	0.23 ± 0.01	0.997	0.25	-0.62 ± 0.02
1a	Н	CH(OH)CF ₃	Н	Н	0.08 ± 0.01	0.863	0.14	-0.34 ± 0.01
1b	Н	COCF ₃	Н	Н	-0.97 ± 0.02	0.999	-0.96	-1.08 ± 0.04
1c	Me	COCF ₃	Н	Me	-1.30 ± 0.02	0.982	-1.27	0.86 ± 0.04
1d	Н	F	Н	Н	-0.21 ± 0.01	0.112	0.74	-3.00 ± 0.19
1e	Me	F	Н	Н	-1.15 ± 0.02	0.666	-0.97	-3.75 ± 0.24
1f	Et	F	Н	Н	-0.75 ± 0.01	0.676	-0.58	-2.08 ± 0.03
1g	<i>n</i> -Pr	F	Н	Н	-0.21 ± 0.05	0.755	-0.08	-0.54 ± 0.05
1h	<i>i</i> -Pr	F	Н	Н	-0.54 ± 0.02	0.74	-0.41	-1.03 ± 0.06
1i	<i>n</i> -Bu	F	Н	Н	0.23 ± 0.02	0.71	0.38	0.99 ± 0.04
1j	Н	F	Н	Me	0.25 ± 0.01	0.166	1.03	-1.91 ± 0.02
1k	Н	F	Me	Н	0.50 ± 0.02	0.154	1.32	-1.88 ± 0.03
11	Н	F	Me	Me	1.07 ± 0.04	0.275	1.64	-0.96 ± 0.00
1m	Me	F	Me	Н	-0.69 ± 0.01	0.899	-0.65	-2.27 ± 0.06
1n	Н	Me	F	Н	-0.003 ± 0	0.971	-0.01	-1.34 ± 0.09
10	Me	Me	F	Н	-0.81 ± 0.01	0.98	-0.80	-2.80 ± 0.04
1p	Н	Н	F	Me	-0.73 ± 0.01	0.95	-0.71	-1.62 ± 0.06
1q	Н	Н	F	Н	-0.92 ± 0.02	0.863	-0.85	-2.67 ± 0.11
1r	Me	Н	F	Н	-1.39 ± 0.01	0.858	-1.32	-3.00 ± 0.06
1s	Et	Н	F	Н	-0.95 ± 0.01	0.876	-0.89	-1.92 ± 0.01
1t	n-Pr	Н	F	Н	-0.63 ± 0.03	0.839	-0.46	-0.61 ± 0.04
1u	i-Pr	Н	F	Н	-1.00 ± 0.02	0.832	-0.92	
1v	Н	F	F	F	-1.31 ± 0.01	0.001	1.70	-1.93 ± 0.17
1w	Н	F	Н	F	-0.36 ± 0.03	0.02	1.35	-3.17 ± 0.51
1x	Н	CF ₃	Н	Н	-0.08 ± 0.02	0.112	0.87	-1.28 ± 0.01
1y	Me	CF ₃	Н	Н	-0.75 ± 0.01	0.5	-0.45	-0.54 ± 0.03
1z	Н	COOMe	Me	Н	-0.32 ± 0.02	0.974	-0.31	0.41 ± 0.07
³ Distribution coefficients were measured using a 10:1 molar ratio of ligand to iron to ensure that the 3:1 neutral complexes formed.								

over the range of 1-12 and computer modeling predictions,²³ 1b and 5-F-HPOs (1n, 1p, 1q) have the same sequence of pK_a values, that is, the lowest pK_a value (<3) is assigned to the protonation of the 4-oxo group, followed by 3-hydroxy group in the range of 6-9 and the p K_a value above 9.5 is assigned to the dissociation of 1-NH (route $pK_{a1a}-pK_{a2a}-pK_{a3a}$ in Scheme 6). However, when fluorine or trifluoromethyl is introduced at the 2-position of 1-unsubstituted HPOs (1d, 1j-1l, 1x), fluorine or trifluoromethyl group dramatically decreases the pK_a value of 1-NH to the range 6.5–7. Correspondingly, the pK_a of 3-hydroxy group is increased due to the influence of 4-O anion. The ionization of this group adopts the route $pK_{a1a}-pK_{a2b}-pK_{a3b}$ (Scheme 6). This result is consistent with the previous report that introduction of 2-F on the pyridine decreases the pK_a value of 1-NH from 5.17 to -0.44^{24} and also with our computer modeling predictions.²³ Furthermore, when two fluorines were introduced at the *ortho*- position of 1-NH (1v and 1w), the p K_a value of 1-NH further drops to <1 and the assignment of the ionization route follows 1-NH to 4-oxo to 3-hydroxy (route $pK_{a1b}-pK_{a2c}-pK_{a3b}$ in Scheme 6).

In comparison with the 1-nonsubstituted analogue 1d, an introduction of methyl group at N1 (1e) dramatically influences the pK_a values of the 4-oxo and 3-hydroxy groups. However, this phenomenon does not occur when fluorine is at C5 position (1n and 1o). On increasing the R₁ chain length

(1e-1i), the pK_{a1} was increased slightly from 2.4 to 2.61 but the pK_{a2} value was unchanged. Again, this trend does not hold when fluorine is at C5 (1r-1t). A shift of the methyl group from C6 to C5 (1j and 1k), is associated with a slight increase of the pK_{a1} value from 1.15 to 1.42 due to the electron donation of methyl group at the ortho- position. Compared to 1q, an introduction of methyl group at C6 (1p) increases both pK_{a1} and pK_{a2} . A further increase was observed when the methyl group was introduced at C2 (1n). In comparison with 2-fluoro-1-alkyl substituted HPOs, the 5-fluoro analogues have lower pK_{a1} values but higher pK_{a2} values (1e vs 1r; 1f vs 1s, 1g vs 1t). This phenomenon can be explained by the fluorine at the *ortho*- position having a stronger influence on the pK_a value than that at the *meta-* position. Similarly, by shifting the fluorine from C5 to C2 in the 1-nonsubstituted F-HPOs (1q vs 1d and 1p vs 1j), the p K_a value of 1-NH is markedly decreased due to the shift of fluorine from *meta*- to *ortho*- position. Surprisingly, the pK_{a1} value, which is assigned to the 4-oxo group, is also decreased (Table 1).

When two or more fluorine atoms are introduced on the pyridinone ring, the pK_a value of 1-NH function is further decreased to <1 (1d to 1w or 1v). In contrast, the pK_a value of the 4-oxo function is increased whereas the pK_a value of the 3-hydroxy function remains in a similar range. Replacement of F-with CF₃- at position 2 generates similar pK_a values (1d to 1x)

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following the same sequence. This is confirmed by the finding that two similar pK_a values were determined when the 3-hydroxy of 1x is protected. The values are 1.4 and 6.1, assigned to 4-oxo and 1-NH, respectively. Similar to the pair of 1d and 1e, an introduction of methyl on N1 dramatically decreases the pK_a value of 3-hydroxy (1y vs 1x).

Determination of Iron(III) Affinity Constants. Stability constants of all F-HPOs ligands were evaluated using an automated spectrophotometric titration system. The value log β_3 was obtained by sum of the logarithms of three stepwise equilibrium constants K_1 , K_2 , and K_3 and are presented in Table 1. The values were found to be related to their pK_a values of both 3-hydroxy and 4-oxo moieties. For N-alkyl F-HPOs, due to their lower pK_a values compared with those of DFP and CP94, the iron stability constants (log β_3) fall correspondingly. This trend also holds for 1b and 1-nonsubstituted 5-F-HPOs (1n, 1p, and 1q) where both chelating related pK_a values (pK_{a1}) and pK_{a2} are less than that of DFP and CP94. The lowest log β_3 value was found to be 26.5 for 1y, where the pK_a values of 3hydroxy and 4-oxo groups are at 7.4 and <0.8, respectively. However, log β_3 values are dramatically increased when pK_a values of the 3-hydroxy group (pK_{a3}) rise to above 10. 1-Nonsubstituted 2-F-HPOs (1d, 1j-1l) and multifluoro substituted HPOs (1v-1x) fell into this group. For example, although the pK, value of the 4-oxo group for 1i is only 1.15, much lower than that of DFP, the increase of pK_a of the 3hydroxy for 1j (11.08 to 9.77 for DFP) leads to a larger log β_3 value than its counterpart (Table 1). The largest log β_3 value in this series is 38.6 for 1w, where two fluorines were attached at C2 and C6, respectively, and pK_a values of the 3-hydroxy and 4oxo groups are 10.6 and 5.7, respectively. However, it is difficult to determine the log β_3 values of 1d, 1j–1l, and 1x precisely due to their 1-NH pK_a values being close to 7. This results in a complication associated with the protonation of the iron complexes, thereby introducing more species in the stoichiometric model. Thus only ranges of the log β_3 and pFe values are provided (Table 1).

By using the pK_a and $\log \beta_3$ values, the pFe^{3+} value, which is defined as the negative logarithm of concentration of free iron(III) at pH 7.4 under conditions $ligand_{[total]} = 10^{-5}$ M and $iron_{[total]} = 10^{-6}$ M, was calculated. pFe^{3+} values of 1-alkyl 2-F-HPOs fall in the same range as that of DFP and CP94, with slightly higher values when the R_1 chain length was increased (1e-1i). However, a dramatic drop of pFe^{3+} value occurs with the 1-unsubstituted counterparts (1d, 1j-11), although the stability constant of 1j-11 are higher than that of DFP. In contrast, there is no marked difference of pFe^{3+} values between 1-alkyl and 1-unsubstituted 5-F-HPOs, with both groups having marginally smaller values than that of DFP.

Determination of Distribution Coefficients. The distribution coefficients $(D_{7,4})$ of the free ligands and their iron(III) complexes between an aqueous phase buffered at pH 7.4 and octanol are presented in Table 2. By replacing hydrogen with fluorine, both distribution coefficients of free ligands and iron complexes fall in a similar range (compare 1m and 1o with DFP), with fluorine located at the second position possessing slightly more lipophilic character. In general, the increase in the alkyl chain length at N1 results in an increase in the $D_{7,4}$ values of both the ligands and the iron(III) complexes (1e-1i and 1r-1t). Similar to our previous report,²⁵ 1-unsubstituted F-HPOs have much higher distribution coefficients compared to their 1-methyl substituted analogues (1d to 1e; 1n to 1o; 1x to 1y). This trend also holds for the

corresponding iron(III) complexes except for the pair 1x and 1y, where the log D_{74} value of 1x is 0.67 higher than that of 1y in contrast to the value of its iron(III) complex, which is 0.74 lower than the counterpart. In comparison with 3-F analogues, 2-F-HPOs are slightly more hydrophobic. For example, the log D_{74} value of 1d is -0.21 whereas that of 1g is -0.92. The same phenomenon is found by comparing 1e with 1r, 1f with 1s, and 1g with 1t. Among these compounds, 1l and 1r possess the highest and the lowest D_{7.4} values, respectively, but do not form the most hydrophobic and hydrophilic iron(III) complexes, respectively. On the other hand, compounds containing similar D_{ligand} values do not necessarily mean that they have similar D_{complex} values (compare 1f to 1y). This observation is different from our previous report that a linear relationship exists between D_{ligand} and D_{complex}^{25} In most cases, iron(III) complexes are more hydrophilic than their corresponding free ligands. However, 1c, which is in the hydrophilic class, possesses relatively hydrophobic property when chelating with iron.

Glucuronidation and Oxidation. One of the main problems for HPOs is that they undergo rapid metabolism in the liver and are glucuronidated at the 3-hydroxyl group, resulting in the formation of the nonchelating 3-O-glucuronide metabolite.⁹ To understand the potential influence of fluorine on the metabolic stability of HPO, several F-HPO chelators were selected for incubation with guinea pig hepatic microsomes. Three were selected which contained a nonalkylated nitrogen atom (1b, 1d, and 1x) and three possessed an *N*alkylated atom (1e, 1f, and 1m). The ratio of glucuronidated HPOs after 1 h incubation to the total added iron chelators was expressed as a percentage conversion. The glucuronidation rates of those F-HPO chelators and deferiprone are presented in Figure 1. 1d and 1x, where the N atom on the ring is



Figure 1. Comparison of the glucuronidation rates of a series of F-HPO chelators with deferiprone incubated for 1 h in guinea pig hepatic microsomes (mean \pm SD) (n = 4 or 5).

unsubstituted, were almost completely metabolized after 1 h. In contrast, **1f** and **1m**, which have similar log P values, were only moderately metabolized under the same conditions, but more rapid than that of deferiprone. Interestingly, substitution of the ethyl group with methyl at N1 (compare **1e** to **1f**) or removing the methyl group at C5 (**1m** to **1e**) leads to increased metabolic stability (Figure 1). Substitution of hydrogen by a methyl group on the heterocyclic nitrogen (compare **1e** to **1d**) dramatically influences metabolic stability. To our surprise, compound **1b**, which has no substitution on N1, was demonstrated to possess a lower glucuronidation rate than deferiprone. This phenomenon may be attributed to the

influence of 2-trifluoroacetyl group, where the presence of a strong intramolecular H-bond between the C=O at position 2 and 3-hydroxy group prevents the rapid metabolism of 3-hydroxyl group.

The two compounds with the lowest rates of glucuronidation (**1b** and **1e**) together with deferiprone were further investigated for phase 1 metabolism. Table 3 shows the relative resistance of

Table 3. Comparison of the Cytochrome P-450 Involved Oxidative Metabolism Rates of Deferiprone, 1b and 1e Incubated 1 h in Guinea Pig Hepatic Microsomes (Mean \pm SD) (n = 5)

substrate	percentage of metabolite (%)
DFP	19.8 ± 1.5
1b	0.6 ± 2.4
1e	3.0 ± 2.0

1b and **1e** to microsomal oxidation as compared to deferiprone. In similar fashion to the glucuronidation result, both F-HPOs were found to possess lower rates of oxidative metabolism than that of deferiprone, indicating potential for improved maintenance of the drug level at a given dose. From this preliminary investigation, it was decided that with the exception of **1b**, *N*-alkylated hydroxypyridinones should be selected for BBB permeability measurements.

Blood–Brain Barrier (BBB) Permeability. Hydroxypyridinones for this study were selected on their basis: their affinity for iron(III) (pFe > 19.0), log *P* value (0 to -1), and with the exception of **1b**, the existence of an *N*-alkyl group. The range of log *P* values was chosen to minimize liver first pass extraction. By adopting these limits, the fluorinated HPOs were reduced to **1b**, **1e–1h**, and **1s–1u**. As **1e–1h** is closely related, we further selected **1f** and **1g** from this subgroup. The brain uptake values at steady-state for these selected F-HPOs and deferiprone are presented in Figure 2. Two F-HPOs (**1g** and **1t**) demonstrated





higher K_p values than deferiprone. If and 1x are also able to permeate the BBB but with less efficiency than deferiprone. Three other F-HPOs, 1s, 1u, and 1b, were found to possess poor BBB permeability. 2-F-HPOs have slightly more lipophilic character when compared to the 5-F analogues, and this may account for the corresponding higher K_p values (1f to 1s and 1g to 1t). Additional methylene groups on the F-HPO also improve brain uptake, probably due to changes in lipophilicity (1g to 1f and 1t to 1s). However, a poor permeability was observed for 1u, an isomer of 1t. This phenomenon may result from its increased hydrophilicity (log *P* value of -0.92 for 1u compared to -0.46 for 1t). The bulky nature of the isopropyl group may be another factor which influences the permeability. Ix is more lipophilic than 1g. However, its K_p value is lower than that of 1g, which indicates that the lipophilicity is not the only determining factor for brain distribution. Although 1b was demonstrated to be metabolically more stable than DFP, it failed to cross BBB. An objective of this study was to identify correlations between the brain distribution and log *P* values of F-HPOs. A clear correlation between the K_p CD values of the F-HPOs and their log *P* values were observed, when the data for 1x is excluded from the study (Figure 3). 1x is the only member



Figure 3. Correlation of KpCD values against log P of F-HPOs (1b, 1f, 1g, 1s, 1t, 1u, and 1x). *y*-Errors are excluded for simplicity. The trendline is drawn excluding the brown circle on the right.

of the group possessing a trifluoromethyl function and despite the relatively high log P value of the free ligand, its ability to permeate the BBB was extremely low.

CONCLUSION

The synthetic route of fluorinated HPOs adopted in this study is different from the conventional method for the HPO synthesis and offers a guideline for the synthesis of other functionalized 3-hydroxypyridin-4-ones where the functional groups are difficult to introduce onto the preformed 3hydroxypyridin-4-one structure. This synthetic approach may be helpful to organic chemists working in the heterocyclic fluorine chemistry field. The physicochemical properties of F-HPOs demonstrate that the introduction of fluorine to 3hydroxypyridin-4-ones markedly influences the pK_a, log β_3 values, and the distribution coefficients of both the free ligands and the iron complexes, when compared with nonfluorinated analogues. Selected F-HPOs were found to possess a range of metabolic stabilities, some being more stable than the clinically used deferiprone. Two of the F-HPOs, namely 1g and 1t, were found to possess superior brain uptake than deferiprone.

EXPERIMENTAL SECTION

General Method. Fluorinated pyridines were purchased from Fluorochem. Reagents were from Sigma-Aldrich and reagent grade quality and were used without further purification. Column chromatography purifications were performed on Merck silica gel 60 (0.04–0.063 mm). ¹H, ¹³C, and ¹⁹F NMR spectra were recorded on a Bruker Avance 400 (400 MHz) NMR spectrometer. Chemical shifts (δ) are reported in ppm downfield from the internal standard

tetramethylsilane (TMS) for ¹H and ¹³C NMR. ESI mass spectra were obtained by infusing samples into an LCQ Deca XP ion trap mass instrument. HRMS were monitored on MicroMass Q-TOF instrument. Purity (\geq 95%) was determined via HPLC analysis.

3-Hydroxy-2-(2,2,2-trifluoro-1-hydroxyethyl)pyridin-4(1*H***)one (1a). To a solution of 1-(3,4-bis(benzyloxy)pyridin-2-yl)-2,2,2trifluoroethanol 3 (1 g) in methanol (15 mL) was added a catalytic amount of Pd/C (5%, 0.1 g) and 1 mL of concentrated HCl. The reaction was hydrogenated at room temperature and 2 atm for 3 h. Then the catalyst was filtered off through Celite, and the residue was concentrated under reduced pressure to afford the title compound as an off-white solid. Recrystallization from EtOH/acetone gave a white solid. Yield 88%. ¹H NMR (DMSO-***d***₆): \delta 8.16 (d,** *J* **= 6.5 Hz, 1H), 7.48 (d,** *J* **= 6.5 Hz, 1H), 5.77 (q,** *J* **= 6.6 Hz, 1H), 4.30 (br s, 3H). ¹⁹F NMR: -72.24 (d,** *J* **= 6.6 Hz, 3F). ¹³C NMR: 63.58 (q,** *J* **= 32 Hz, CHOH), 112.06 (CH-5), 123.90 (q,** *J* **= 282 Hz, CF₃), 132.99 (C-2), 135.80 (CH-6), 143.38 (C-3), 161.63 (CO-4). HRMS: calcd for C₇H₇NO₃F₃ (M + 1)⁺, 210.0378; found, 210.0382.**

General Procedure for the Preparation of 1b–q and 1v–1z. Methyl or ethyl protecting 3,4-dihydroxypyridine or 3-hydroxy-1-alkyl-pyridin-4-one was dissolved into CH_2Cl_2 (20 mL) and flushed with nitrogen at -5 °C. Boron trichloride or boron tribromide (1 M in CH_2Cl_2 , 8 mL) was slowly added, and the reaction mixture was stirred at room temperature for 20 h. The excess BCl_3/BBr_3 was eliminated at the end of the reaction by the addition of methanol (10 mL) and left to stir for another half an hour. After removal of the solvents under reduced pressure, the residues were purified by recrystallization to afford white solids.

3-Hydroxy-2-(2,2,2-trifluoroacetyl)pyridin-4(1H)-one Hydrochloride (**1b**). Yield: 70%. ¹H NMR (DMSO- d_6): δ 8.17 (d, J = 6.4 Hz, 1H), 7.46 (d, J = 6.4 Hz, 1H), 4.08 (br s, 2H). ¹⁹F NMR: -78.54 (s, 3F). ¹³C NMR: 113.37 (CH-5), 121.70 (q, J = 155 Hz, CF₃), 131.37 (C-2), 134.53 (CH-6), 145.45 (C-3), 163.31 (CO-4), 168.68 (COCF₃). HRMS: calcd for C₇H₅NO₃F₃ (M + 1)⁺, 208.0222; found, 208.0219.

2-(2,2,2-Trifluoroacetyl)-3-hydroxy-1,6-dimethylpyridin-4(1H)one Hydrochloride (1c). Yield: 65%. ¹H NMR (DMSO- d_6) δ 8.73 (brs, 1H, OH), 7.28 (s, 1H, C-5H), 4.18 (s, 3H, NCH₃), 2.62 (CH₃). ¹⁹F NMR: -81.77 (d, *J* = 7.4 Hz). ¹³C NMR: 21.13 (s, CH₃), 41.48 (s, *J* = 20 Hz, NCH₃), 113.58 (s, CH-5), 122.20 (d, *J* = 306 Hz, CF₃), 134.54 (s, C-2), 145.17 (s, C-3), 150.91 (s, C-6), 160.93 (s, C-4), 214.00 (s, COCF₃). HRMS: calcd for C₉H₉NO₃F₃ (M + 1)⁺, 236.0535; found, 236.0539.

2-Fluoro-3-hydroxypyridin-4(1H)-one Hydrobromide (1d). Yield: 60%. ¹H NMR (CD₃OD): δ 7.41 (d, J = 5.5 Hz, 1H, C-6H), 6.72 (d, J = 5.5 Hz, 1H, C-5H)), 4.94 (brs, 2H, OH and NH). ¹⁹F NMR (CD₃OD): -85.48 (s). ¹³C NMR (CD₃OD): 111.67 (d, J = 3 Hz, C-5), 129.35 (d, J = 29 Hz, C-3), 137.17 (d, J = 16 Hz, C-6), 155.75 (d, J = 228 Hz, C-2), 157.58 (d, J = 9 Hz, C-4). HRMS: calcd for C₅H₃NO₂F (M + 1)⁺, 130.0304; found, 130.0314.

2-Fluoro-3-hydroxy-1-methylpyridin-4(1H)-one Hydrobromide (1e). Yield: 66%. ¹H NMR (CD₃OD): δ 8.06 (dd, J = 5.1, 7.1 Hz, 1H, C-6H), 7.13 (dd, J = 0.8 7.1 Hz, 1H, C-5H), 4.96 (brs, 1H, OH), 4.09 (d, J = 3.8 Hz, 3H, CH₃). ¹⁹F NMR (CD₃OD): -102.49 (s). ¹³C NMR (CD₃OD): 40.70 (d, J = 6 Hz, CH₃), 111.39 (s, C-5), 133.03 (d, J = 12 Hz, C-3), 136.10 (d, J = 5 Hz, C-6), 152.32 (d, J = 265 Hz, C-2), 165.05 (d, J = 10 Hz, C-4). HRMS: calcd for C₆H₇NO₂F (M + 1)⁺, 144.0461; found, 144.0460.

1-*E*thyl-2-fluoro-3-hydroxypyridin-4(1*H*)-one Hydrobromide (**1f**). Yield: 59%. ¹H NMR (DMSO-*d*₆): δ 8.17 (dd, *J* = 5.3, 7.1 Hz, 1H, C-6H), 7.11 (d, *J* = 7.1 Hz, 1H, C-5H), 4.37 (dq, *J* = 2.5, 7.2 Hz, 2H, CH₂), 1.41 (t, *J* = 7.2 Hz, 3H, CH₃). ¹⁹F NMR (DMSO-*d*₆): -106.84 (s). ¹³C NMR (DMSO-*d*₆): 14.81 (s, CH₃), 48.69 (d, *J* = 4 Hz, CH₂), 110.50 (s, C-5), 131.15 (d, *J* = 12 Hz, C-3), 133.60 (d, *J* = 5 Hz, C-6), 149.98 (d, *J* = 264 Hz, C-2), 163.89 (d, *J* = 11 Hz, C-4). HRMS: calcd for C₇H₉NO₂F (M + 1)⁺, 158.0617; found, 158.0622.

2-Fluoro-3-hydroxy-1-propylpyridin-4(1H)-one Hydrobromide (1g). Yield: 70%. ¹H NMR (DMSO- d_6): δ 8.14 (dd, J = 5.3, 7.1 Hz, 1H, C-6H), 7.09 (d, J = 7.1 Hz, 1H, C-5H), 4.30 (t, J = 7.1 Hz, 2H, CH₂), 1.85–1.78 (m, 2H, CH₂), 0.89 (t, J = 7.3 Hz, 3H, CH₃). ¹⁹F

NMR: -107.45 (s). ¹³C NMR: 10.31 (s, CH₃), 22.54 (s, CH₂), 54.31 (d, J = 2 Hz, CH₂), 110.47 (s, CH-5), 131.19 (d, J = 11 Hz, C-3), 134.02 (d, J = 4 Hz, CH-6), 149.88 (d, J = 264 Hz, C-2), 164.42 (d, J = 10 Hz, C-4). HRMS: calcd for C₈H₁₁NO₂F (M + 1)⁺, 172.0774; found, 172.0777.

2-Fluoro-3-hydroxy-1-isopropylpyridin-4(1H)-one Hydrobromide (1h). Yield: 60%. ¹H NMR (DMSO- d_6): δ 8.25 (dd, J = 5.1, 7.2 Hz, 1H, C-6H), 7.13 (d, J = 7.2 Hz, 1H, C-5H), 5.13–4.93 (m, 1H, CH), 4.70 (brs, OH), 1.53 (d, J = 6.7 Hz, 6H, 2CH₃). ¹⁹F NMR: -105.59. ¹³C NMR: 21.32 (s, CH₃), 55.77 (d, J = 5 Hz, CH), 110.64 (s, C-5H), 131.09 (d, J = 13 Hz, C-3), 143.76 (d, J = 22 Hz, CH-6), 149.90 (d, J = 272 Hz, C-2), 163.01 (d, J = 11 Hz, C-4). HRMS: calcd for C₈H₁₁NO₂F (M + 1)⁺, 172.0774; found, 172.0774.

1-Butyl-2-fluoro-3-hydroxypyridin-4(1H)-one Hydrobromide (1i). Yield: 64%. ¹H NMR (DMSO- d_6): δ 8.18 (d, J = 5.4, 7.0 Hz, 1H, C-6H), 7.12 (d, J = 7.1 Hz, 1H, C-5H), 4.34 (t, J = 7.1 Hz, 2H, CH₂), 1.80–1.72 (m, 2H, CH₂), 1.35–1.26 (m, 2H, CH₂), 0.91 (t, J = 7.4 Hz, 3H, CH₃). ¹⁹F NMR: -107.11 (s). ¹³C NMR: 13.30 (s, CH₃), 18.79 (s, CH₂), 31.06 (s, CH₂), 52.81 (d, J = 3 Hz, CH₂), 110.46 (s, CH-5), 131.20 (d, J = 11 Hz, C-3), 134.01 (d, J = 4 Hz, CH-6), 149.91 (d, J = 264 Hz, C-2), 164.25 (d, J = 10 Hz, C-4). HRMS: calcd for C₉H₁₃NO₂F (M + 1)⁺, 186.0930; found, 186.0925.

2-*Fluoro-3-hydroxy-6-methyl-pyridin-4(1H)-one Hydrobromide* (1j). Yield: 78%. ¹H NMR (DMSO-*d*₆): δ 6.61 (s, 1H, C-5H), 5.99 (brs, OH and NH), 2.21 (s, 3H, CH₃). ¹⁹F NMR (DMSO-*d*₆): -90.71 (s). ¹³C NMR (DMSO-*d*₆): 22.23 (s, CH₃), 109.50 (d, *J* = 3 Hz, C-5), 125.04 (d, *J* = 29 Hz, C-3), 144.52 (d, *J* = 13 Hz, C-6), 152.92 (d, *J* = 227 Hz, C-2), 156.26 (d, *J* = 8 Hz, C-4). HRMS: calcd for C₆H₇NO₂F (M + 1)⁺, 144.0461; found, 144.0463.

2-*Fluoro-3-hydroxy-5-methyl-pyridin-4(1H)-one* Hydrobromide (**1k**). Yield: 74%. ¹H NMR (DMSO- d_6): δ 7.59 (brs, OH and NH), 7.36 (s, 1H, C-6H), 2.07 (s, 3H, CH₃). ¹⁹F NMR (DMSO- d_6): -91.60 (s). ¹³C NMR (DMSO- d_6): 12.53 (s, CH₃), 119.84 (d, *J* = 3 Hz, C-5), 126.56 (d, *J* = 29 Hz, C-3), 135.54 (d, *J* = 16 Hz, C-6), 152.69 (d, *J* = 224 Hz, C-2), 154.12 (C-4). HRMS: calcd for C₆H₇NO₂F (M + 1)⁺, 144.0461; found, 144.0478.

2-Fluoro-3-hydroxy-5,6-dimethyl-pyridin-4(1H)-one Hydrobromide (1I). Yield: 75%. ¹H NMR (DMSO- d_6): δ 6.23 (brs, OH and NH), 2.23 (s, 3H, 6-CH₃), 2.02 (s, 3H, 5-CH₃). ¹⁹F NMR (DMSO- d_6): -94.63 (s). ¹³C NMR (DMSO- d_6): 11.03 (s, 5-CH₃), 20.43 (s, 6-CH₃), 117.10 (d, *J* = 4 Hz, C-5), 124.59 (d, *J* = 29 Hz, C-3), 142.63 (d, *J* = 13 Hz, C-6), 150.60 (d, *J* = 227 Hz, C-2), 155.10 (d, *J* = 8 Hz, C-4). HRMS: calcd for C₇H₉NO₂F (M + 1)⁺, 158.0617; found, 158.0609.

2-*Fluoro-3-hydroxy-1,5-dimethyl-pyridin-4(1H)-one Hydrobromide (1m).* Yield: 79%. ¹H NMR (DMSO-*d*₆): δ 8.19 (d, J = 5.4Hz, 1H, C-6H), 6.15 (brs, 1H, OH), 3.95 (d, J = 3.7 Hz, 3H, 1-CH₃), 2.14 (s, 3H, 5-CH₃). ¹⁹F NMR (DMSO-*d*₆): -105.50 (s). ¹³C NMR (DMSO-*d*₆): 12.60 (s, 5-CH₃), 39.48 (d, J = 5 Hz, 1-CH₃), 120.41 (s, C-5), 129.51 (d, J = 13 Hz, C-3), 133.68 (d, J = 5 Hz, C-6), 149.57 (d, J = 263 Hz, C-2), 162.73 (d, J = 9 Hz, C-4). HRMS: calcd for C₇H₉NO₂F (M + 1)⁺, 158.0617; found, 158.0627.

5-Fluoro-3-hydroxy-2-methylpyridin-4(1H)-one Hydrobromide (1n). Yield: 69%. ¹H NMR (DMSO- d_6): δ 8.55 (d, *J* = 5.0 Hz, 1H, C-6H), 4.54 (brs, OH and NH), 2.54 (s, 3H, CH₃). ¹⁹F NMR (DMSO- d_6): -148.82 (s). ¹³C NMR (DMSO- d_6): 14.04 (s, CH₃), 121.88 (d, *J* = 32 Hz, C-6), 137.31 (s, C-2), 144.11 (d, *J* = 7 Hz, C-3), 148.73 (d, *J* = 238 Hz, C-5), 150.48 (d, *J* = 12 Hz, C-4). HRMS: calcd for C₆H₇NO₂F (M + 1)⁺, 144.0461; found, 144.0468.

5-Fluoro-3-hydroxy-1,2-dimethyl-pyridin-4(1H)-one Hydrobromide (10). Yield: 62%. ¹H NMR (DMSO- d_6): δ 8.72 (d, J = 6.1 Hz, 1H, C-6H), 4.89 (brs, OH), 3.99 (s, 3H, 1-CH₃), 2.47 (s, 3H, 2-CH₃). ¹⁹F NMR (DMSO- d_6): -149.61 (d, J = 6.1 Hz). ¹³C NMR (DMSO- d_6): 12.70 (s, 2-CH₃), 43.91 (s, 1-CH₃), 127.24 (d, J = 35 Hz, C-6), 139.28 (s, C-2), 144.58 (d, J = 7 Hz, C-3), 147.60 (d, J = 237 Hz, C-5), 149.80 (d, J = 13 Hz, C-4). HRMS: calcd for C₇H₉NO₂F (M + 1)⁺, 158.0617; found, 158.0617.

3-Fluoro-5-hydroxy-2-methylpyridin-4(1H)-one Hydrobromide (1p). Yield: 72%. ¹H NMR (DMSO- d_6): δ 8.04 (d, J = 0.5 Hz, 1H, C-6H), 4.30 (brs, OH and NH), 2.64 (d, J = 2.8 Hz, 3H, CH₃). ¹⁹F NMR (DMSO- d_6): -146.32 (s). ¹³C NMR (DMSO- d_6): 12.67 (s,

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CH₃), 123.02 (s, C-6), 134.77 (d, J = 27 Hz, C-2), 145.25 (d, J = 7 Hz, C-5), 147.32 (d, J = 238 Hz, C-3), 150.60 (d, J = 11 Hz, C-4). HRMS: calcd for C₆H₇NO₂F (M + 1)⁺, 144.0461; found, 144.0468.

3-Fluoro-5-hydroxypyridin-4(1H)-one Hydrobromide (1q). Yield: 65%. ¹H NMR (DMSO-*d*₆): δ 8.64 (dd, *J* = 1.0, 4.9 Hz, 1H, C-2H), 8.12 (s, 1H, C-6H). ¹⁹F NMR (DMSO-*d*₆): -146.96 (s). ¹³C NMR (DMSO-*d*₆): 123.91 (d, *J* = 33 Hz, C-2), 124.66 (s, C-6), 146.93 (d, *J* = 8 Hz, C-5), 149.50 (d, *J* = 240 Hz, C-3), 151.58 (d, *J* = 12 Hz, C-4). HRMS: calcd for C₅H₅NO₂F (M + 1)⁺, 130.0304; found, 130.0301.

2,3,6-Trifluoro-5-hydroxypyridin-4(1H)-one Hydrochloride (1v). Yyield: 72%. ¹⁹F NMR (DMSO- d_6): δ –93.30 (dd, J = 18.8, 22.6 Hz, 1F), -100.84 (dd, J = 18.8, 26.3 Hz, 1F), -166.00 (dd, J = 22.6, 26.3 Hz, 1F). ¹³C NMR: 127.06 (d, J = 31 Hz, C-3), 135.07 (dd, J = 18, 241 Hz, C-5), 140.40 (dd, J = 14, 224 Hz, C-2), 144.28 (dd, J = 15, 230 Hz, C-6), 147.78 (s, C-4). HRMS: calcd for C₅H₃NO₂F₃ (M + 1)⁺, 166.0116; found, 166.0139.

2,6-Difluoro-3-hydroxypyridin-4(1H)-one Hydrobromide (1w). Yield: 67%. ¹H NMR (DMSO- d_6): δ 10.81 (br s, 1H), 8.93 (br s, 1H), 6.31 (d, J = 0.5 Hz, 1H). ¹⁹F NMR: -79.37 (d, J = 15.7 Hz, 1F), -90.59 (d, J = 15.7 Hz, 1F). ¹³C NMR: 96.13 (dd, J = 4.5, 40.5 Hz, CH-5), 127.60 (dd, J = 6.5, 27.5 Hz, C-3), 152.80 (dd, J = 19, 232 Hz, C-2), 155.39 (dd, J = 18, 231 Hz, C-6), 161.74 (dd, J = 8, 12 Hz, CO-4). HRMS: calcd for $C_5H_4NO_2F_2$ (M + 1)⁺, 148.0210; found, 148.0236.

2-(*Trifluoromethyl*)-3-hydroxypyridin-4(1H)-one Hydrochloride (1**x**). Yield: 76%. ¹H NMR (DMSO- d_6): δ 8.52 (br s), 7.99 (d, J = 5.6 Hz, 1H), 7.29 (d, J = 5.6 Hz, 1H). ¹⁹F NMR: -63.49 (s, 3F). ¹³C NMR: 113.72 (s, CH-5), 121.58 (q, J = 273 Hz, CF₃), 129.34 (q, J = 33 Hz, C-2), 138.83 (s, CH-6), 143.55 (s, C-3), 157.63 (s, CO-4). HRMS: calcd for C₆H₅NO₂F₃ (M + 1)⁺, 180.0274; found, 180.0275.

3-Hydroxy-1-methyl-2-trifluoromethyl-1H-pyridin-4-one Hydrobromide (**1y**). Yield: 52%. ¹H NMR (DMSO- d_6): δ 8.84 (d, J = 6.8 Hz, 1H), 7.68 (d, J = 6.7 Hz, 1H), 4.24 (d, J = 1.1 Hz, 3H). ¹⁹F NMR: -57.37 (s, 3F). ¹³C NMR: 45.77 (d, J = 4 Hz, CH₃), 108.09 (s, CH-5), 119.35 (q, J = 37 Hz, C-2), 119.76 (q, J = 274 Hz, CF₃), 146.98 (s, CH-6), 150.09 (s, C-3), 164.85 (s, CO-4). HRMS: calcd for C₇H₇NO₂F₃ (M + 1)⁺, 194.0459; found, 194.0443.

Methyl 3-Hydroxy-5-methyl-4-oxo-1,4-dihydropyridine-2-carboxylate Hydrobromide (1z). Yield: 78%. ¹H NMR (DMSO- d_6): δ 8.15 (s, 1H), 5.65 (brs), 4.07 (s, 3H), 2.28 (s, 3H). ¹³C NMR: 13.42 (CH₃), 53.70 (OCH₃), 121.36 (C-5), 124.12 (C-2), 136.87 (C-6), 146.79 (C-3), 162.32 (CO-4), 164.06 (2-COO). HRMS: calcd for C₈H₁₀NO₄ (M + 1)⁺, 184.0610; found, 184.0606.

General Procedure for Preparation of 1r-u. A solution of 30 (5 mmol) in acetone (30 mL) was added alkyl iodide (10 mmol), and the mixture was heated at 60 °C overnight. The solvent was then evaporated and the residue was recrystallized to afford white solids.

3-*Fluoro-5-hydroxy-1-methylpyridin-4(1H)-one* (1*r*). Yield: 86%. ¹H NMR (DMSO-*d*₆): δ 8.68 (d, *J* = 5.2 Hz, 1H, C-2H), 8.08 (s, 1H, C-6H), 5.20 (brs, 1H, OH), 4.04 (s, 3H, CH₃). ¹⁹F NMR (DMSO-*d*₆): -147.39 (s). ¹³C NMR (DMSO-*d*₆): 45.95 (s, CH₃), 127.81 (d, *J* = 35 Hz, C-2), 128.53 (s, C-6), 146.91 (d, *J* = 8 Hz, C-5), 148.96 (d, *J* = 238 Hz, C-3), 150.94 (d, *J* = 12 Hz, C-4). HRMS: calcd for C₆H₇NO₂F (M + 1)⁺, 144.0461; found, 144.0465.

1-*E*thyl-3-fluoro-5-hydroxypyridin-4(1H)-one (**1s**). Yield: 82%. ¹H NMR (DMSO-*d*₆): δ 8.04 (d, *J* = 6.0 Hz, 1H, C-2H), 7.57 (s, 1H, C-6H), 4.62 (brs, OH), 3.93 (q, *J* = 7.1 Hz, 2H, CH₂), 1.32 (t, *J* = 7.1 Hz, 3H, CH₃). ¹⁹F NMR (DMSO-*d*₆): -156.02 (d, *J* = 6.0 Hz). ¹³C NMR (DMSO-*d*₆): 15.98 (s, CH₃), 51.51 (s, CH₂), 121.69 (s, C-6), 124.36 (d, *J* = 35 Hz, C-2), 149.35 (d, *J* = 15 Hz, C-5), 150.63 (d, *J* = 233 Hz, C-3), 160.66 (d, *J* = 13 Hz, C-4). HRMS: calcd for C₇H₉NO₂F (M + 1)⁺, 158.0617; found, 158.0609.

3-Fluoro-5-hydroxy-1-propylpyridin-4(1H)-one (1t). Yield: 70%; refluxed for 3 days. ¹H NMR (DMSO- d_6): δ 8.03 (dd, J = 2.1, 7.1 Hz, 1H, C-2H), 7.53 (d, J = 2.0 Hz, 1H, C-6H), 3.85 (t, J = 7.1 Hz, 2H, CH₂), 1.77–1.68 (m, 2H, CH₂), 0.81 (t, J = 7.4 Hz, 3H, CH₃). ¹⁹F NMR: -156.36 (d, J = 7.4 Hz). ¹³C NMR: 10.29 (s, CH₃), 23.46 (s, CH₂), 57.62 (s, CH₂), 121.95 (s, CH-6), 124.73 (d, J = 35 Hz, CH-2), 149.23 (d, J = 13 Hz, C-5), 150.51 (d, J = 233 Hz, C-3), 160.75 (d, J = 13 Hz, C-4). HRMS: calcd for $C_8H_{11}NO_2F$ (M + 1)⁺, 172.0774; found, 172.0769.

3-Fluoro-5-hydroxy-1-isopropylpyridin-4(1H)-one (1u). Yield: 65%; refluxed in 2-iodopropane only. ¹H NMR (DMSO- d_6): δ 8.45 (dd, *J* = 2.0, 6.7 Hz, 1H, C-2H), 7.91 (d, *J* = 1.9 Hz, C-6H), 4.53–4.46 (m, 1H, CH), 1.44 (d, *J* = 6.7 Hz, 6H, 2CH₃). ¹⁹F NMR: –150.74 (s). ¹³C NMR: 22.06 (s, CH₃), 60.13 (s, CH), 123.01 (s, C-6), 123.65 (d, *J* = 34 Hz, C-2), 148.24 (d, *J* = 10 Hz, C-5), 150.11 (d, *J* = 237 Hz, C-3), 155.79 (d, *J* = 13 Hz, C-4). HRMS: calcd for C₈H₁₁NO₂F (M + 1)⁺, 172.0774; found, 172.0768.

ASSOCIATED CONTENT

S Supporting Information

Details of the synthesis, physicochemical property, metabolism and blood-brain barrier studies, the relationship between log D_{ligand} and log $D_{\text{iron complex}}$ of the fluorinated HPOs. This material is available free of charge via the Internet at http:// pubs.acs.org.

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Notes

The authors declare no competing financial interest.

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ABBREVIATIONS USED

DFP, deferiprone; HPO, 3-hydroxypyridin-4-one; CP94, 1,2diethyl-3-hydroxypyridin-4-one; LDA, lithium diisopropylamide; LTMP, lithium 2,2,6,6-tetramethylpiperidine; USB, unimetal superbase; BBB, blood-brain barrier

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